Narrative Review

Diagnostic salivary biomarkers in traumatic brain injury: narrative review

Shahideh Rostami^{® 1}, Fatemeh Heidarzadeh^{® 1}, Sara Ashourzadeh Fallah^{® 1}, Seyed Alireza Rahimi^{® 1}, Maryam Mehrizi^{® 1}, Fatemeh Sadeghi^{® 1}, MohammadHossein Shahrabadi^{® 1}, Marziyeh Hajizadeh^{® 1}, Jennifer Swann ^{® 2}, Alireza Jalali Farahani^{® 3}, Seyed Morteza Hosseiniara^{® 1*}

¹Medical Student, Faculty of Medicine, Tehran Medical Sciences, Islamic Azad University, Tehran, Iran

² Professor, Department of Biological Sciences, 111 Research Drive, Lehigh University, Bethlehem, PA 18015, USA

³ Professor of Cardiac Anesthesia, School of Medicine, Atherosclerosis Research Center, Baqiyatallah University of Medical sciences, Tehran, Iran

* Corresponding author: Seyed Morteza Hosseiniara. Faculty of Medicine, Tehran Medical Sciences, Islamic Azad University, Tehran, Iran Email: hosseiniara89@gmail.com

Received: 30 January 2022 Revised: 5 February 2022 Accepted: 18 February 2022 e-Published: 1 March 2022

Abstract

Traumatic brain injury (TBI) is a common cause of disability and mortality worldwide. TBI is an acquired brain injury that may be open (penetrating) or closed (non-penetrating) and is be categorized as mild, moderate, or severe, depending on the clinical presentation. Accurate diagnosis at the earliest stages can significantly affect patient discomfort, prognosis, therapeutic intervention, survival rates and recurrence. Whereas traditional CT and MRI techniques for diagnosis are dominant in clinical situations, a promising direction for clinical diagnosis is the use of fluid biomarkers like blood, CSF, urine, and saliva. Fluid biomarkers that may track these injuries and inflammatory processes have been explored for their potential to provide objective measures in TBI assessment. At present, there are limited clinical guidelines available regarding the use of fluid biomarkers in TBI.

In recent years, saliva has received significant attention as a biomarker for TBI in clinical practice due to the non-invasive accessibility, cost-effective collection, and consistent relationship with serum. This review examines the utility of saliva biomarkers such as S100B, noncoding RNAs (ncRNAs), extracellular vesicles (EVs), miRNAs levels, microtubule-associated protein tau, alpha-amylase, cortisol, and oxidative stress in TBI.

The study highlights the current state of salivary diagnostics, future aspirations, and their potential as the preferred route of TBI detection. The newly developed techniques for salivary analysis of these molecules may help to improve outcomes for TBI through rapid detection current unavailable with serum samples. Future studies via salivary biomarkers will help establish consistent strategies for early diagnosis of TBI and improve treatment outcomes of TBI patients.

Keywords: Saliva, Traumatic Brain Injury, Head trauma, Biomarker.

Introduction

Globally, traumatic brain injury (TBI), is the greatest single contributor to disability and death of all traumarelated injuries. The accurate identification and diagnosis of TBI is the first step toward providing appropriate clinical care. However, accurate clinical identification of patients with TBI is complicated by variations in the criteria used for diagnosis.¹ There are no conclusive biologic tools to detect TBI or to track brain recovery, diagnosis and management largely consist of patientreported symptoms and subjective clinical assessment. There is increasing interest in employing saliva samples as a source of biomarkers for TBI in clinical practice due to the non-invasive accessibility, cost-effective collection, and consistent relationship with serum.² New methods for saliva handling, analysis, and biomarker discovery offer strong promise for the components of saliva as biomarkers for TBI. These methodological advances have identified

Copyright© 2022. This open-access article is published under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License which permits Share (copy and redistribute the material in any medium or format) and Adapt (remix, transform, and build upon the material) under the Attribution-NonCommercial terms. Downloaded from: https://www.nclinmed.com/

new candidate biomarkers of TBI, and present the feasibility of their use for diagnostics and prognostics of TBI.³ If successfully validated, saliva biomarkers of TBI will break new ground by improving clinical management of TBI and advancing TBI treatments.

Traumatic brain injury

Damage to the brain after trauma is referred to as traumatic brain injury (TBI). TBI may be blunt, nonpenetrating, penetrating, or the result of a blast. The resulting neuropathology consists of a primary injury that is a direct consequence of the traumatic insult and a secondary injury that results from a cascade of molecular and cellular events triggered by the primary injury and that leads to cell death, axonal injury, and inflammation. The annual incidence of TBI has been estimated to be 27 to 69 million, worldwide. Many survivors live with significant disabilities, resulting in a major socioeconomic burden. The symptoms of a TBI can be mild, moderate, or severe, depending on the extent of damage to the brain. Mild trauma may induce brief changes in mental state or consciousness. Severe trauma may induce prolonged periods of unconsciousness, coma, or even death.^{4,5}

A computed tomography scan (CT scan) is the gold standard for the radiological assessment of a TBI patient. A CT scan is easy to perform and an excellent test for detecting the presence of blood and fractures, critical markers in medical trauma cases. Plain x-rays of the skull are recommended by some as a way to evaluate patients with only mild neurological dysfunction. Magnetic resonance imaging (MRI) is not commonly used for acute head injury since it takes longer to perform an MRI than a CT. Because it is difficult to transport an acutely injured patient from the emergency room to an MRI scanner, the use of MRI is impractical. Besides these accurate tools, the assessment of biomarkers in biofluids like blood, CSF, urine, and saliva have received attention and present some valid results.⁶

Despite progress in preventive and therapeutic strategies, delay in TBI diagnosis remains one of the major causes of high morbidity and mortality. The prevalence of TBI is rising and leads to an increase in the burden of the socioeconomic and health system, so rapid and accurate diagnosis of TBI is necessary.

Saliva as a source of biomarkers

Human saliva is a clear and slightly acidic heterogeneous biofluid (pH 6.0 to 7.0) composed of water (99%), proteins (0.3%), and minerals (0.2%). On average, individual saliva secretion can vary from 0.3 to 0.7 ml of saliva per minute, producing a range of 1 to 1.5 liters per day. Saliva assists as in a variety of functions including tasting, swallowing, and digestion of food. Saliva also plays a role in fat deposition and serves as a protective barrier against pathogens.⁷

Saliva is produced in the salivary glands by acinous cells, and released into the oral cavity through a series of ducts. The parotid, submandibular, and sublingual, create more than 90% of total saliva, with the smaller glands, (lip, buccal, lingual, and palate) providing the rest.^{7,8}

Saliva is collected and analyzed as unstimulated whole saliva, unstimulated saliva from specific glandular pairs (e.g. parotid or submandibular and sublingual pairs), or stimulated saliva from specific pairs of glands. Whole saliva that is present in the oral cavity for 24-hour periods is characterized as "unstimulated whole saliva" and is normally associated with precise clinical conditions when compared to stimulated saliva, because the substances used to prime the flow may affect saliva composition. Unstimulated saliva is collected from an individual's mouth by passively allowing it to flow into a container, or saliva is held in the mouth for a specific period and spit into a container. This method of collecting is the "gold standard" for obtaining many saliva components.⁹

Saliva contains several growth factors including EGF, FGF, NGF, and TGF- α , that are essential for the regeneration of the oral and esophageal mucosa. Some antibacterial and antifungal components are also found in saliva, such as lysozyme, immunoglobulins, and lactoferrin, that prevent the progression of bacterial infection and tooth decay. An important proteolytic enzyme, α -amylase, is secreted by saliva. Some of these components may serve as diagnostic biomarkers, that can be accurately analyzed using specific and sensitive immunological and biochemical techniques such as RIA, ELISA, and chromatography.¹⁰

Recent technological advances in the processing and evaluation of salivary components have yielded reliable results increasing the characterization of this biological resource as a relatively safer, cheaper, and less invasive measure than traditional samples such as blood. Collecting blood carries potential risks to individuals, including transient discomfort, bruising, infection at the site of the vessel, and anemia (if large volumes or vulnerable individuals are required). Saliva samples can have several advantages over blood for routine TBI testing, including their safe and easy collection.¹¹

According to the National Institutes of Health (NIH), a biomarker is an objective indicator of the measurement and evaluation of natural biological processes, pathogenic processes, or drug responses to therapeutic interventions whose concentration, structure, function, or inactivity are correlated with the onset, progression, or even regression of a particular disorder or as a result of the body's response to it. Thus, biomarkers act as valuable and attractive tools in the diagnosis, risk assessment, diagnosis, prognosis, and monitoring of the disease.¹²

The use of salivary biomarkers as new tools for the diagnosis of TBI are emerging. Recent studies that have examined the use of biomarkers in the saliva, including S100B, noncoding RNAs (ncRNAs), extracellular vesicles (EVs), miRNAs levels, microtubule-associated protein tau, alpha-amylase, cortisol, and oxidative stress are described below.

Salivary S100B

Protein S100B is a member of the S100 protein family and comprises the largest subset of Ca²⁺ EF-hand-binding proteins. S100B is mainly synthesized by astrocytes in the human brain and secreted from glial cells into the extracellular space, where it exhibits cytokine-like functions, and modulates long-term synaptic plasticity.¹³

The structure of the S100B protein, provided by the X-Ray technique, is an octagon of four homodimeric units arranged as two tetramers in a tight array. In Escherichia coli, several other recombinant types of S100B, including tetrameric, hexameric, and octameric, have been identified. Related studies have shown that the binding of the S100B tetrameric structure to RAGE receptors (recipients of advanced glycation end products) occurs with greater affinity than its dimeric structure. In this regard, Tetramer S100B activates cell growth more strongly than dimer S100B and increases cell survival.¹⁴

Although the highest amount of S100B has been measured in glial cells of the central nervous system its persistence and extracellular function lead to the detection of this protein in non-CNS cells such as skeletal myofibers, myoblasts, adipose tissue, and melanocytes, and other tissues. Due to the presence of this biomarker in the systemic circulation, it has the ability to be present, examined, and measured in biological body fluids such as blood, urine, saliva, and amniotic fluid.¹⁵

The S100B levels in body fluids, including blood and saliva increase after TBI, heavy exercise, and neurological diseases. Therefore, an increase in the level of S100B can be considered as a sign of nerve cell damage. A study of 15 adult patients with suspected TBI (mean age= 47 years, range 18–79) and 15 control subjects (mean age= 33 years, range 23–53) found salivary S100B levels were 3.9 fold higher than blood S100B, regardless of the presence of pathology. The salivary level of this protein was as accurate in differentiating TBI patients from control subjects as serum levels. These preliminary results suggest that salivary S100B measurements could be a reliable alternative to serum S100B in the diagnosis of TBI. Further studies with a large sample size are needed to confirm these findings.¹¹

Early detection of blood-brain barrier damage and intracranial hemorrhage within minutes is critical to prevent permanent brain disability after TBI. This detection requires measurement of the S100B protein level with some 100% validity. To measure serum levels, plasma samples are separated by centrifugation within 2 hours of collection, the serum is collected and stored at -70 ° C. Serum levels of S100B protein are determined using an ELISA kit. The same kit can be used to directly detect the protein S100B in saliva (without centrifugation, separation, or storage), the time saved is an important benefit of saliva over plasma. Utilizing salivary samples of this protein in cases of TBI has other potential benefits including non-invasive sampling and lack of infection risk.16

Salivary noncoding RNAs (ncRNAs)

Non-coding RNA (ncRNA), discovered in 1950, is RNA that is not translated into a protein. Instead, ncRNAs act as guides directing proteins to specific target sites and play important roles in differentiation, epigenetic regulation, transcription regulation, and post-transcription regulation.

ncRNAs are widely distributed in various tissues, although some ncRNAs have tissue-specific expression.

Intracellular localization of ncRNAs can be found in a wide range of intracellular components such as the cytoplasm, nucleus, or both. The number of ncRNAs in the human genome is unknown; however, recent transcriptomic and bioinformatics studies suggest that this classification includes thousands of sequences. ncRNAs are classified by length [Figure-1]. These functionally important types of ncRNAs are including tRNA, rRNA, microRNAs, siRNAs, piRNAs, snoRNAs, snRNAs, exRNAs, and scaRNAs. There are several epigenetic changes, including DNA methylation and histone modification, and changes in ncRNA levels that may occur as a result of TBI.17-19

One of the most important features of ncRNAs is their high stability, especially when exposed to exosomes and apoptotic bodies. ncRNA is detectable in body fluids like blood and saliva suggesting their potential usage as biomarkers for screening, diagnosis, prognosis, and response to treatment and evaluation of treatment.²⁰

In the UK Men's Rugby Union, physicians used ncRNAs as salivary biomarkers for the assessment of head injuries in athletes.²¹ A study on the hippocampus of rats showed that the expression level of circRNA, which is a type of ncRNA, changed significantly after TBI. These results suggest that altered circRNA expression patterns in the rat hippocampus after TBI may play important roles in post-TBI physiological and pathological processes. These findings may provide not only a new direction for studying the molecular mechanisms underlying TBI but also a new possibility for the treatment of TBI by modulating circRNAs.²² In another study, RNA sequencing was performed on 505 saliva samples obtained longitudinally from 112 individuals (8-24-years-old) with TBI. ncRNA biomarkers show promise for tracking recovery from TBI, and for predicting who will have prolonged symptoms. Salivary ncRNA levels represent a non-invasive and biological measurement that could aid in the accurate, early diagnosis of TBI, improving clinical outcomes for

Salivary MicroRNAs

MicroRNAs (miRNAs) are small, endogenous, noncoding molecules with a length of 19-42 nucleotides that were discovered in 1993. miRNA genes can be very different in position in the genome. We have 2 separate classes of miRNAs: One group is encoded by proteinencoding transcription introns and the next group is encoded by exons. miRNAs are evolutionary RNAs that act as regulators of post-transcriptional expression and affect the translation of proteins throughout the body. miRNA primarily inhibits or modifies the production of a protein produced by binding to complementary target sequences in mRNA and interfering with translation machines. In addition to suppressing translation, the binding of miRNA to mRNA causes the uptake and association of mRNA decay factors and leads to mRNA instability, degradation, and consequently reduced expression levels.24,25

miRNA is found in every human tissue and biofluid. The CNS contains the highest concentration and highest diversity of miRNAs. It is estimated that 70% of all miRNAs are expressed in the brain, spinal cord, or peripheral nerves. These molecules are resistant to RNAse degradation, have the ability to cross the BBB, and are transported through protected extracellular space in exosomes and microvesicles, allowing them to be easily measured in biofluids, including serum, cerebrospinal fluid, and saliva.²⁶

Due to their abundance, stability in pH fluctuations, resistance to enzymatic degradation, and their key role in transcriptional regulation, miRNAs are potentially ideal biomarkers for the diagnosis, identification, and classification of cancers and other diseases, including neurodegenerative diseases, diabetes, and TBI patients. These molecules are critical to the function of neurons and regulate gene expression in response to brain damage.²⁷

Saliva can receive exosomal miRNAs directly from the cranial nerves in the mouth and throat, therefore responds faster than blood miRNAs that must pass through the BBB. Salivary miRNAs, on the other hand, reflect the secondary neuroplasticity response, which explains their high diagnostic accuracy.²⁷

Diagnostic salivary biomarkers in traumatic brain injury: narrative review

Salivary Extracellular vesicles

Most studies have examined miRNA levels in peripheral blood, and a few recent studies have identified saliva as a new biofluid. Hicks et al., suggest that these miRNAs could predict TBI status in individuals, and salivary miRNA levels were associated with the severity of mental symptoms.²⁸ In another study, saliva samples were collected at multiple time points, both pre-and post-fight from 50 amateurs mixed martial arts fighters. A subset of salivary miRNAs showed robust utility at predicting TBI likelihood and demonstrated quantitative associations with head impacts as well as cognitive and balance measures.²⁹

Extracellular vesicles (EVs) are bilayer vesicles of phospholipid membranes that are secreted by different cells. These particles contain biological molecules such as mRNA, miRNA, DNA, proteins, and lipids. EVs were initially described as a way for homeostasis and biological waste disposal. Today, the important and influential role of EVs in intracellular communication is very evident not only in normal functions but also in disease conditions such as cancer, autoimmune diseases, cardiovascular and respiratory failure as well as trauma, and tissue damage.³⁰

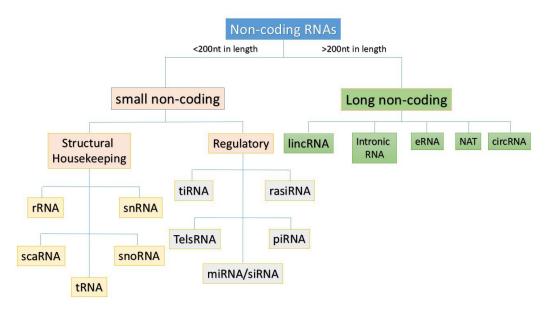


Figure 1. Classification of non-coding RNAs

EVs may affect bone marrow stromal regeneration, target cell function, angiogenesis, progression, and metastasis of blood malignancies by inducing gene expression in target cells. EVs mediate communication between cells by affecting target cells and regulate physiological and pathological processes.^{31,32}

The recent classification of EVs is based on biophysical characteristics such as size, cellular origin, molecular content, and biogenesis. According to these criteria, EVs are categories to exosomes, microvesicles, and apoptotic bodies, and oncosomes.³³

These EVs contain the genomic and proteomic content of the stem cell, can be used as potential biomarkers, and are available from biological fluids such as serum, urine, and saliva.³⁴ Accurate and fast diagnosis of TBI is vital for patients. In recent years, there has been a growing interest in EVs that contain a variety of biomarkers and as an indicator of the status of target cells. In fact, EVs can be considered as a biopsy platform in the field of early diagnosis and treatment. Studies have shown that EVs released by damaged cells to biological fluids can be used as potential biomarkers to diagnose and evaluate the severity of TBI.

EVs move between the brain and the oral cavity and can be isolated from salivary specimens as a non-invasive TBI biomarker that may highlight severe changes or neuropathology.^{35,36}

In a study of 19 subjects with 7 healthy controls, 6 patients diagnosed with concussion injury from an outpatient concussion clinic, and 6 patients with TBI who received treatment in the emergency department. Real-

time PCR analysis of salivary extracellular vesicles in participants showed significant changes in the expression of 15 inflammation-related genes. These findings indicate that inflammation biomarkers can be used for the diagnosis of TBI and the evaluation of disease severity.³⁵ Also, findings from another study conducted on 8 mixed martial arts (MMA) fighters and 7 controls, suggest that salivary EVs may be used as a biomarker in the acute period following TBI to identify the severity of the injury severity and to elucidate pathophysiological processes involved in TBI.³⁷ Considering the consistent results in several studies, as well as the non-invasiveness and relatively fast sampling from saliva, it can be mentioned that salivary EVs can be used as a diagnostic biomarker to detect TBI.

Microtubule-associated protein tau

Microtubule-associated protein tau (MAPT) is a multifunctional intracellular protein that belongs to the MT-associated protein family. MAPT is predominantly expressed in neurons and is highly enriched in the axonal regions. In the human brain, MAPT presents high immunoreactivity in non-myelinated axons of cortical interneurons located in the grey matter. MAPT has been recognized as a biomarker of axonal disruption and the release of MAPT into blood and CSF serves as an indication of neurotrauma. Elevated levels of MAPT have been observed in mild and severe TBI.³⁸⁻⁴⁰

Although the role of MAPT as a biomarker in the case of TBI has been investigated by examination of CSF and blood obtained during a forensic autopsy, less is known concerning MAPT liberation and occurrence in other biofluids like saliva. MAPT levels were significantly elevated in saliva of those in a study group that was selected following neuropathological examination compared to the control group.⁴² The elevated MAPT concentration levels in saliva were predictive of the axonal injury even in cases where the head injury was not considered to be the direct cause of death and thus were undiagnosed and omitted during the regular forensic autopsy.⁴³ In another study, the elevated salivary MAPT concentration levels using ELISA were recorded in cases of TBI in population-based autopsy screening, as a biomarker for axonal injury.⁴²

MAPT release in saliva is provided by two distinct mechanisms. Secretion of saliva is mediated by the salivary principal glands such as sublingual, submandibular, and parotid that are innervated through parasympathetic components of the cranial nerves VII (facial), IX (glossopharyngeal), and their associated submandibular and otic ganglia.44 This close anatomical relation of salivary glands with the nervous system supports the hypothesis that MAPT is directly released from their innervation. Alternatively, MAPT could pass from systemic blood circulation into the saliva through the blood-saliva barrier which exchanges kinetics of molecules that remain largely unaltered in the event of head trauma.⁴⁵ Thus, elevated concentration levels of MAPT in saliva should be considered as a potential biomarker for TBI in postmortem examination.

Salivary alpha-amylase

Salivary alpha-amylase (sAA) is a digestive enzyme produced by salivary acinar cells that break down alpha-1 to 4 glycosidic bonds in starch and related carbohydrates to produce glucose, maltose, maltotriose, and dextrin. During the past years, sAA has presented as a valid and reliable biomarker for autonomic nervous system (ANS) activity in stress research.⁴⁶ It has also been observed as a non-invasive biomarker for sympathetic nervous system (SNS) activity.

In a prospective cohort study, sAA was elevated in children 13-15 yrs old after TBI and the levels of sAA predicted specific post-traumatic stress symptoms in children.47 In another study, Salivary alpha-amylase increased in response to trauma reflection in 20 women (mean age 23.6 +/- 5.8 years) with a history of trauma exposure. Salivary alpha-amylase might serve as a more reliable marker of trauma-related reactivity to negative affective information, and also as a marker of hypervigilance in the absence of threatening information.48 As sAA is one of the most abundant components of saliva, it may prove to be a reliable biomarker in the diagnosis of TBI and stress.

Salivary cortisol

Cortisol is a low molecular weight lipid steroid hormone that is produced and secreted by the cortical part of the adrenal glands. Hormone production is regulated by the hypothalamus in the brain and pituitary gland (HPA axis). Most cortisol in the blood (up to 95%) is protein-bound and only a small percentage is "free" and biologically active. Free cortisol is present in saliva, too. Studies have shown that salivary cortisol can be kept stable for at least 5-7 days at Room temperature.^{49,50}

In a study, relative to a healthy non-injured comparison group, TBI-injured children (ages 8-12 years) had higher cortisol levels following vehicular accidents.⁴⁷ In another study, the most common abnormalities included cortisol elevation in 34 adult TBI patients, following weaning from mechanical ventilation.⁵¹ Conversely, another study salivary cortisol was not increased in response to the trauma reminder in 20 women with a history of trauma exposure.⁴⁸

Because a salivary stress marker is widely accepted, low levels of cortisol in saliva were evaluated as a non-invasive measure to modulate stress responses. Apilux et al's development of a lateral current immunoassay device using a cortisol-BSA conjugate containing gold-labeled gold nanoparticles in a silver amplification system holds promise as an effective tool to detect cortisol associated with salivary stress.⁵²

The assessment of salivary cortisol as a diagnostic biomarker in TBI is recommended. TBI may cause anxiety and stress in patients, and the positive correlation between stress and cortisol levels support its potential as a biomarker in TBI.⁵³

Oxidative Stress

Reactive oxygen species (ROS) and reactive nitrogen species (RNS) are produced during natural physiological processes. ROS and RNS are highly reactive molecules that can damage key cell components. Under physiological conditions, the endogenous immune system is able to prevent the formation and aids in the removal of these harmful molecules protecting tissues against oxidative damage. Oxidative stress plays a crucial role in pathology. Typically, cells contain several antioxidants to counteract the damaging effects of oxidative chemicals, and a healthy biological balance between ROS and antioxidants must be maintained to prevent oxidative damage to cells and tissues.Any oxidative imbalance leads to the accumulation of oxidants causing oxidative damage to cells.53,54

In either case, changes in biomarkers can indicate the severity of the abnormality or the extent of the damage. The overall change in oxidative or antioxidant markers is a reliable indicator, and individual interpretation of each oxidative stress marker will be useful for a more accurate diagnosis and for understanding the overall trend.^{53,54}

There is very little literature on the importance of antioxidants in TBI. Brain ascorbic acid decreased in TBI due to experimental explosion and was associated with decreased Glutathione and thiol protein and increased oxidative markers. Glutathione S-Transferases (GST) is an enzyme that exhibits glutathione peroxidase activity. Natural changes in GST expression affect neuronal degradation after experimental TBI and confirm the importance of lipid peroxidation as an important pathophysiological mechanism in TBI.^{56,57}

Oxidative stress markers, including ROS and their mediators, have been evaluated in saliva samples. Salivary biomarkers have been shown to play a profound role in the diagnosis of many diseases or stressful conditions, including TBI.^{58,59}

New insights into saliva cells as the main source of salivary biomarkers have prompted the development of new assay methods for standard measurement of biomarkers in fluid as well as cells. Digital Immunohistochemistry (IHC) and Western Blot (WB) for quantitative measurement of SRP 100 biomarker have been used for rapid detection.⁶⁰

Other Biomarkers

Several other biomarkers including glial fibrillar acidic protein (GFAP), ubiquitin carboxy-terminal hydrolase L1 (UCH-L1), neuron-specific enolase (NSE), neurofilament light protein (NFL), beta-amyloid protein, lysozyme, immunoglobulins, lactoferrin, and others in saliva that may serve as diagnostic biomarkers in TBI patients need further studies.

Conclusions

More than half of all adults will be exposed to a traumatic event at some point in their lives, yet we do not yet have reliable biomarkers to help predict who experiences trauma-related symptoms in response to these events. In

Rostami et al

this regard, it seems that several reliable, less expensive, and easily attainable biomarkers for TBI in saliva provide hope for early detection that may save lives.

This review of the literature has identified saliva biomarkers with the highest discriminative abilities as determined by operating characteristics in diagnostic situations in TBI. Saliva shows a strong promise for biomarkers of TBI based on new methods for saliva handling, analysis, and biomarker discovery.

The top performers in each biomarker may provide insight into pathogenic mechanisms of TBI that most influence the measured endpoint. Nonetheless, many challenges remain before these biomarkers can be incorporated into clinical practice. In particular, it remains unclear whether a large panel of biomarkers in addition to clinical assessment will be sufficient to stratify patients into categories of TBI before more specific biomarker assessments are applied.

With the refinement and validation of saliva biomarkers, we have the potential to use saliva biomarkers as a convenient and cost-effective surrogate for currently used imaging modalities for the evaluation of TBI in the clinical setting.

Acknowledgment

None.

Competing interests

The authors declared no potential conflict of interests with respect to the research, authorship, and/or publication of this article.

Abbreviations

Traumatic brain injury: TBI; Computed tomography scan: CT scan; Magnetic resonance imaging: MRI; National Institutes of Health: NIH; Noncoding RNAs: ncRNAs; Extracellular vesicles: EVs; MicroRNAs: miRNAs; Microtubule-associated protein tau: MAPT; Mixed martial arts: MMA; Glutathione S-Transferases: GST: Immunohistochemistry: IHC; Western Blot: WB; Reactive oxygen species: ROS; Reactive nitrogen species: RNS; Salivary alpha-amylase: sAA; Autonomic nervous system: ANS; Sympathetic nervous system: SNS.

Authors' contributions

All authors pass the four criteria for authorship contribution based on the International Committee of Medical Journal Editors (ICMJE) recommendations. All authors read and approved the final manuscript. All authors take responsibility for the integrity of the data and the accuracy of the data analysis.

Funding

The authors received no financial funding or support for the research.

Role of the funding source

None.

Availability of data and materials

The data used in this study are available from the corresponding author on request.

Ethics approval and consent to participate

None.

Consent for publication

By submitting this document, the authors declare their consent for the final accepted version of the manuscript to be considered for publication.

References

- Rubiano AM, Carney N, Chesnut R, Puyana JC. Global neurotrauma research challenges and opportunities. Nature. 2015; 527:S193-S197. doi:10.1038/nature16035 PMid:26580327
- Cheng Y, Pereira M, Raukar N, Reagan JL, Queseneberry M, Goldberg L, et al. Potential biomarkers to detect traumatic brain injury by the profiling of salivary extracellular vesicles. J Cell Physiol. 2019; 234(8):14377-88. doi:10.1002/jcp.28139 PMid:30644102 PMCid:PMC6478516
- Monteleone MC, Billi SC, Viale L, Catoira NP, Frasch AC, Brocco MA. Search of brain-enriched proteins in salivary extracellular vesicles for their use as mental disease biomarkers: a pilot study of the neuronal glycoprotein M6a. J Affect Disord Rep. 2020;1:100003. doi:10.1016/j.jadr.2020.100003
- Courtney A, Courtney M. The complexity of biomechanics causing primary blast-induced traumatic brain injury: a review of potential mechanisms. Front Neurol. 2015;6:221. doi:10.3389/fneur.2015.00221 PMid:26539158 PMCid:PMC4609847
- 5. Helmick KM, Spells CA, Malik SZ, Davies CA, Marion DW, Hinds SR. Traumatic brain injury in the US military: epidemiology and key clinical and research programs. Brain

Imaging Behav. 2015; 9(3):358-66. doi:10.1007/s11682-015-9399-z PMid:25972118

- Buttram SD, Garcia-Filion P, Miller J, Youssfi M, Brown SD, Dalton HJ, et al. Computed tomography vs magnetic resonance imaging for identifying acute lesions in pediatric traumatic brain injury. Hosp Pediatr. 2015;5(2):79-84. doi:10.1542/hpeds.2014-0094 PMid:25646200
- de Almeida PD, Gregio AM, Machado MA, De Lima AA, Azevedo LR. Saliva composition and functions: a comprehensive review. J Contemp Dent Pract. 2008;9(3):72-80. doi:10.5005/jcdp-9-3-72 PMid:18335122
- Tiwari M. Science behind human saliva. J Nat Sci Biol Med. 2011; 2(1):53. doi:10.4103/0976-9668.82322 PMid:22470235 PMCid:PMC3312700
- Munro CL, Grap MJ, Jablonski R, Boyle A. Oral health measurement in nursing research: state of the science. Biol Res Nurs. 2006;8(1):35-42. doi:10.1177/1099800406289343 PMid:167666627 PMCid:PMC2213421
- Altin KT, Topcuoglu N, Duman G, Unsal M, Celik A, Kuvvetli SS, et al. Antibacterial effects of saliva substitutes containing lysozyme or lactoferrin against Streptococcus mutans. Arch Oral Biol. 2021:105183. doi:10.1016/j.archoralbio.2021.105183 PMid:34091207
- Janigro D, Kawata K, Silverman E, Marchi N, Diaz-Arrastia R. Is salivary S100B a biomarker of traumatic brain injury? A pilot study. Front Neurol. 2020;11:528. doi:10.3389/fneur.2020.00528 PMid:32595592 PMCid:PMC7303321
- Menetski JP, Hoffmann SC, Cush SS, Kamphaus TN, Austin CP, Herrling PL, et al. The Foundation for the National Institutes of Health Biomarkers Consortium: past accomplishments and new strategic direction. Clin Pharmacol Ther. 2019; 105 (4):829-43. doi:10.1002/cpt.1362 PMid:30648736 PMCid:PMC6593617
- Michetti F, Bruschettini M, Frigiola A, Abella R, Giamberti A, Marchese N, et al. Saliva S100B in professional sportsmen: high levels at resting conditions and increased after vigorous physical activity. Clin Biochem. 2011;44(2-3):245-7. doi:10.1016/j.clinbiochem.2010.10.007 PMid:20970414
- Ostendorp T, Leclerc E, Galichet A, Koch M, Demling N, Weigle B, et al. Structural and functional insights into RAGE activation by multimeric \$100B. EMBO J. 2007;26(16):3868-78. doi:10.1038/sj.emboj.7601805 PMid:17660747 PMCid:PMC1952220
- Koh SX, Lee JK. S100B as a marker for brain damage and bloodbrain barrier disruption following exercise. Sports Med. 2014; 44 (3):369-85. doi:10.1007/s40279-013-0119-9 PMid:24194479
- Michetti F, D'Ambrosi N, Toesca A, Puglisi MA, Serrano A, Marchese E, et al. The S100B story: from biomarker to active factor in neural injury. J Neurochem. 2019;148(2):168-87. doi:10.1111/jnc.14574PMid:30144068
- Sone M, Hayashi T, Tarui H, Agata K, Takeichi M, Nakagawa S. The mRNA-like noncoding RNA Gomafucons titutes a novel nuclear domain in a subset of neurons. J Cell Sci. 2007;120 (15): 2498-506 doi:10.1242/jcs.009357 PMid:17623775
- 18. Tano K, Mizuno R, Okada T, Rakwal R, Shibato J, Masuo Y, et al. MALAT-1 enhances cell motility of lungadenocarcinoma cells by influencing the expression of motility-related genes. FEBS Lett. 2010;584(22):4575-80 doi:10.1016/j.febslet.2010.10.008 PMid:20937273

- Mateen BA, Hill CS, Biddie SC, Menon DK. DNA methylation: basic biology and application to traumatic brain injury. J Neurotrauma. 2017;34(16):2379-88. doi:10.1089/neu.2017.5007 PMid:28482743
- DiStefano JK. The emerging role of long noncoding RNAs in human disease. Dis Gene Identif. 2018:91-110. doi:10.1007/978-1-4939-7471-9_6 PMid:29423795
- Fuller CW, Fuller GW, Kemp SP, Raftery M. Evaluation of World Rugby's concussion management process: results from Rugby World Cup 2015. Br J Sports Med. 2017;51(1):64-9. doi:10.1136/bjsports-2016-096461 PMid:27587799
- 22. Xie BS, Wang YQ, Lin Y, Zhao CC, Mao Q, Feng JF, et al. Circular RNA expression profiles alter significantly after traumatic brain injury in rats. J Neurotrauma. 2018;35(14):1659-66. doi:10.1089/neu.2017.5468 PMid:29357736
- 23. Fedorchak G, Rangnekar A, Onks C, Loeffert AC, Loeffert J, Olympia RP, et al. Saliva RNA biomarkers predict concussion duration and detect symptom recovery: a comparison with balance and cognitive testing. J Neurol. 2021:1-3. doi:10.1007/s00415-021-10566-x PMid:34028616 PMCid:PMC8505318
- 24. Lee RC, Feinbaum RL, Ambros V. The C. elegans heterochronic gene lin-4 encodes small RNAs with antisense complementarity to lin-14. Cell. 1993;75(5):843-54. doi:10.1016/0092-8674(93)90529-Y
- 25. Rodriguez A, Griffiths-Jones S, Ashurst JL, Bradley A. Identification of mammalian microRNA host genes and transcription units. Genome Res. 2004;14(10a):1902-10. doi:10.1101/gr.2722704 PMid:15364901 PMCid:PMC524413
- 26. Adlakha YK, Saini N. Brain microRNAs and insights into biological functions and therapeutic potential of brain enriched miRNA-128. Mol Cancer. 2014;13(1):1-8. doi:10.1186/1476-4598-13-33 PMid:24555688 PMCid:PMC3936914
- Wittmann J, Jäck HM. Serum microRNAs as powerful cancer biomarkers. Biochim Biophys Acta Bioenerg. 2010;1806(2):200-7. doi:10.1016/j.bbcan.2010.07.002 PMid:20637263
- 28. Hicks SD, Johnson J, Carney MC, Bramley H, Olympia RP, Loeffert AC, et al. Overlapping microRNA expression in saliva and cerebrospinal fluid accurately identifies pediatric traumatic brain injury. J Neurotrauma. 2018;35(1):64-72. doi:10.1089/neu.2017.5111 PMid:28762893 PMCid:PMC7227420
- 29. LaRocca D, Barns S, Hicks SD, Brindle A, Williams J, Uhlig R, et al. Comparison of serum and saliva miRNAs for identification and characterization of mTBI in adult mixed martial arts fighters. PloS one. 2019;14(1):e0207785. doi:10.1371/journal.pone.0207785 PMid:30601825 PMCid:PMC6314626
- Camussi G, Deregibus MC, Bruno S, Cantaluppi V, Biancone L. Exosomes/microvesicles as a mechanism of cell-to-cell communication. Kidney Int. 2010;78(9):838-48. doi:10.1038/ki.2010.278 PMid:20703216
- Santiago-Dieppa DR, Steinberg J, Gonda D, Cheung VJ, Carter BS, Chen CC. Extracellular vesicles as a platform for 'liquid biopsy'in glioblastoma patients. Expert Rev Mol Diagn. 2014; 14 (7):819-25. doi:10.1586/14737159.2014.943193 PMid:25136839 PMCid:PMC4436244
- 32. Hu T, Wolfram J, Srivastava S. Extracellular vesicles in cancer

Rostami et al

detection: hopes and hypes. Trends Cancer. 2020. doi:10.1016/j.trecan.2020.09.003 PMid:33008796

- Dabrowska S, Andrzejewska A, Janowski M, Lukomska B. Immunomodulatory and Regenerative Effects of Mesenchymal Stem Cells and Extracellular Vesicles: Therapeutic Outlook for Inflammatory and Degenerative Diseases. Front Immunol. 2021; 11:3809. doi:10.3389/fimmu.2020.591065 PMid:33613514 PMCid:PMC7893976
- 34. Massa M, Croce S, Campanelli R, Abbà C, Lenta E, Valsecchi C, Avanzini MA. Clinical applications of mesenchymal stem/stromal cell derived extracellular vesicles: therapeutic potential of an acellular product. Diagnostics. 2020;10(12):999. doi:10.3390/diagnostics10120999 PMid:33255416 PMCid:PMC7760121
- 35. Cheng Y, Pereira M, Raukar NP, Reagan JL, Quesenberry M, Goldberg L, et al. Inflammation-related gene expression profiles of salivary extracellular vesicles in patients with head trauma. Neural Regen Res. 2020;15(4):676. doi:10.4103/1673-5374.266924 PMid:31638091 PMCid:PMC6975135
- 36. Chiabotto G, Gai C, Deregibus MC, Camussi G. Salivary extracellular vesicle-associated exRNA as cancer biomarker. Cancers. 2019;11(7):891. doi:10.3390/cancers11070891 PMid:31247906 PMCid:PMC6679099
- 37. Matuk R, Pereira M, Baird J, Dooner M, Cheng Y, Wen S, et al. The role of salivary vesicles as a potential inflammatory biomarker to detect traumatic brain injury in mixed martial artists. Sci Rep. 2021 ;11(1). doi:10.1038/s41598-021-87180-4 PMid:33854105 PMCid:PMC8047010
- 38. Spillantini MG, Goedert M. Tau pathology and neurodegeneration. Lancet Neurol. 2013;12(6):609-22. doi:10.1016/S1474-4422(13)70090-5
- 39. Rubenstein R, Chang B, Davies P, Wagner AK, Robertson CS, Wang KK. A novel, ultrasensitive assay for tau: potential for assessing traumatic brain injury in tissues and biofluids. J Neurotrauma. 2015;32(5):342-52. doi:10.1089/neu.2014.3548 PMid:25177776 PMCid:PMC4348038
- 40. Olczak M, Niderla-Bielińska J, Kwiatkowska M, Samojłowicz D, Tarka S, Wierzba-Bobrowicz T. Tau protein (MAPT) as a possible biochemical marker of traumatic brain injury in postmortem examination. Forensic Sci Int. 2017;280:1-7. doi:10.1016/j.forsciint.2017.09.008 PMid:28942078
- Blomberg M, Jensen M, Basun H, Lannfelt L, Wahlund LO. Cerebrospinal fluid tau levels increase with age in healthy individuals. Dement Geriatr Cogn Disord. 2001;12(2):127-32. doi:10.1159/000051246 PMid:11173885
- 42. Olczak M, Poniatowski ŁA, Niderla-Bielińska J, Kwiatkowska M, Chutorański D, Tarka S, et al. Concentration of microtubule associated protein tau (MAPT) in urine and saliva as a potential biomarker of traumatic brain injury in relationship with bloodbrain barrier disruption in postmortem examination. Forensic Sci Int. 2019;301:28-36. doi:10.1016/j.forsciint.2019.05.010 PMid:31128406
- 43. Jashnani KD, Kale SA, Rupani AB. Vitreous humor: biochemical constituents in estimation of postmortem interval. J Forensic Sci. 2010;55(6):1523-7. doi:10.1111/j.1556-4029.2010.01501.x PMid:20666922
- 44. Farah R, Haraty H, Salame Z, Fares Y, Ojcius DM, Sadier NS. Salivary biomarkers for the diagnosis and monitoring of

neurological diseases. Biomed J. 2018;41(2):63-87. doi:10.1016/j.bj.2018.03.004 PMid:29866603 PMCid:PMC6138769

- 45. Dadas A, Janigro D. The role and diagnostic significance of cellular barriers after concussive head trauma. Concussion. 2018;3(1):CNC53. doi:10.2217/cnc-2017-0019 PMid:30202595 PMCid:PMC6093276
- 46. Ali N, Nater UM. Salivary alpha-amylase as a biomarker of stress in behavioral medicine. Int J Behav Med. 2020;27(3):337-42. doi:10.1007/s12529-019-09843-x PMid:31900867 PMCid:PMC7250801
- 47. Ewing-Cobbs L, Prasad MR, Cox Jr CS, Granger DA, Duque G, Swank PR. Altered stress system reactivity after pediatric injury: Relation with post-traumatic stress symptoms. Psychoneuroendocrinology. 2017;84:66-75. doi:10.1016/j.psyneuen.2017.06.003 PMid:28667938 PMCid:PMC5555029
- 48. Yoon SA, Weierich MR. Salivary biomarkers of neural hypervigilance in trauma-exposed women.
 Psychoneuroendocrinology. 2016;63:17-25. doi:10.1016/j.psyneuen.2015.09.007 PMid:26398002 PMCid:PMC4695293
- 49. Kirschbaum C, Hellhammer DH. Salivary cortisol. Encyclopedia of stress. 2000;3(379-383).
- Garde AH, Hansen ÅM. Long-term stability of salivary cortisol. Scandinavian J Clin Lab Investig. 2005; 65(5):433-6. doi:10.1080/00365510510025773 PMid:16081365
- 51. Dimopoulou I, Tsagarakis S, Theodorakopoulou M, Douka E, Zervou M, Kouyialis AT, et al. Endocrine abnormalities in critical care patients with moderate-to-severe head trauma: incidence, pattern and predisposing factors. Intensive Care Med. 2004;30(6):1051-7. doi:10.1007/s00134-004-2257-x PMid:15069597
- 52. Apilux A, Rengpipat S, Suwanjang W, Chailapakul O. Development of competitive lateral flow immunoassay coupled with silver enhancement for simple and sensitive salivary cortisol detection. EXCLI J. 2018;17:1198.
- Ebrecht M, Hextall J, Kirtley LG, Taylor A, Dyson M, Weinman J. Perceived stress and cortisol levels predict speed of wound healing in healthy male adults. Psychoneuroendocrinology. 2004; 29 (6): 798-809. doi:10.1016/S0306-4530(03)00144-6
- 54. Hanafy KA, Selim MH. Antioxidant strategies in neurocritical care. Neurotherapeutics. 2012;9(1):44-55. doi:10.1007/s13311-011-0085-6 PMid:22135010 PMCid:PMC3271156
- 55. Yoshikawa T, Naito Y. What is oxidative stress?. Japan med Assoc J. 2002;45(7):271-6.
- 56. Tavazzi B, Signoretti S, Lazzarino G, Amorini AM, Delfini R, Cimatti M, et al. Cerebral oxidative stress and depression of energy metabolism correlate with severity of diffuse brain injury in rats. Neurosurgery. 2005;56(3):582-9. doi:10.1227/01.NEU.0000156715.04900.E6 PMid:15730584
- 57. Al Nimer F, Ström M, Lindblom R, Aeinehband S, Bellander BM, Nyengaard JR, et al. Naturally occurring variation in the glutathione-S-transferase 4 gene determines neurodegeneration after traumatic brain injury. Antioxid Redox Signal. 2013; 18(7): 784-94. doi:10.1089/ars.2011.4440 PMid:22881716 PMCid:PMC3555113
- 58. Gümüş P, Emingil G, Öztürk VÖ, Belibasakis GN, Bostanci N.

Oxidative stress markers in saliva and periodontal disease status: modulation during pregnancy and postpartum. BMC Infect Dis. 2015;15(1):1-9. doi:10.1186/s12879-015-1003-z PMid:26152310 PMCid:PMC4495776

- 59. Khurshid Z, Zafar MS, Khan RS, Najeeb S, Slowey PD, Rehman IU. Role of salivary biomarkers in oral cancer detection. Adv Clin Chem. 2018;86:23-70. doi:10.1016/bs.acc.2018.05.002 PMid:30144841
- 60. Walt DR, Blicharz TM, Hayman RB, Rissin DM, Bowden M, Siqueira WL, et al. Microsensor arrays for saliva diagnostics. Ann N Y Acad Sci. 2007;1098(1):389. doi:10.1196/annals.1384.031 PMid:17435144 PMCid:PMC7168095
- 61. McMahon PJ, Panczykowski DM, Yue JK, Puccio AM, Inoue T, Sorani MD, et al. Measurement of the glial fibrillary acidic protein and its breakdown products GFAP-BDP biomarker for the detection of traumatic brain injury compared to computed tomography and magnetic resonance imaging. J Neurotrauma. 2015;32 (8):527-33. doi:10.1089/neu.2014.3635 PMid:25264814 PMCid:PMC4394160
- 62. Mondello S, Palmio J, Streeter J, Hayes RL, Peltola J, Jeromin A. Ubiquitin carboxy-terminal hydrolase L1 (UCH-L1) is increased in cerebrospinal fluid and plasma of patients after epileptic seizure. BMC Neurol. 2012;12(1):1-7. doi:10.1186/1471-2377-12-85 PMid:22931063 PMCid:PMC3500207
- 63. Mayo S, Benito-León J, Peña-Bautista C, Baquero M, Cháfer-Pericás C. Recent Evidence in Epigenomics and Proteomics Biomarkers for Early and Minimally Invasive Diagnosis of Alzheimer's and Parkinson's Diseases. Curr Neuropharmacol. 2021; 19 (8): 1273-303. doi:10.2174/1570159X19666201223154009 PMid:33357195 PMCid:PMC8719284

Cite this article as:

Rostami S, Heidarzadeh F, Ashourzadeh Fallah S, Rahimi SA, Mehrizi M, Sadeghi F, et al. Diagnostic salivary biomarkers in traumatic brain injury: narrative review. Novel Clin Med. 2022; 1(2):59-69. doi: 10.22034/NCM.2022.327121.1009